

Research article

Open Access

Parallel expression evolution of oxidative stress-related genes in fiber from wild and domesticated diploid and polyploid cotton (*Gossypium*)

Bhupendra Chaudhary^{1,2}, Ran Hovav¹, Lex Flagel¹, Ron Mittler^{3,4} and Jonathan F Wendel*¹

Address: ¹Department of Ecology, Evolution and Organismal Biology, Iowa State University, Ames, Iowa 50011, USA, ²School of Biotechnology, Gautam Buddha University, Greater Noida, 201 308 UP, India, ³Department of Biochemistry and Molecular Biology, MS200, University of Nevada, Reno NV 89557, USA and ⁴Department of Plant Sciences, Hebrew University of Jerusalem, Givat Ram, Jerusalem 91904, Israel

Email: Bhupendra Chaudhary - bhupendra@gbu.ac.in; Ran Hovav - ranh@volcani.agri.gov.il; Lex Flagel - flagel@iastate.edu; Ron Mittler - ronmittler@gmail.com; Jonathan F Wendel* - jfw@iastate.edu

* Corresponding author

Published: 17 August 2009

Received: 2 February 2009

BMC Genomics 2009, 10:378 doi:10.1186/1471-2164-10-378

Accepted: 17 August 2009

This article is available from: <http://www.biomedcentral.com/1471-2164/10/378>

© 2009 Chaudhary et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Reactive oxygen species (ROS) play a prominent role in signal transduction and cellular homeostasis in plants. However, imbalances between generation and elimination of ROS can give rise to oxidative stress in growing cells. Because ROS are important to cell growth, ROS modulation could be responsive to natural or human-mediated selection pressure in plants. To study the evolution of oxidative stress related genes in a single plant cell, we conducted comparative expression profiling analyses of the elongated seed trichomes ("fibers") of cotton (*Gossypium*), using a phylogenetic approach.

Results: We measured expression changes during diploid progenitor species divergence, allopolyploid formation and parallel domestication of diploid and allopolyploid species, using a microarray platform that interrogates 42,429 unigenes. The distribution of differentially expressed genes in progenitor diploid species revealed significant up-regulation of ROS scavenging and potential signaling processes in domesticated *G. arboreum*. Similarly, in two independently domesticated allopolyploid species (*G. barbadense* and *G. hirsutum*) antioxidant genes were substantially up-regulated in comparison to antecedent wild forms. In contrast, analyses of three wild allopolyploid species indicate that genomic merger and ancient allopolyploid formation had no significant influences on regulation of ROS related genes. Remarkably, many of the ROS-related processes diagnosed as possible targets of selection were shared among diploid and allopolyploid cultigens, but involved different sets of antioxidant genes.

Conclusion: Our data suggests that parallel human selection for enhanced fiber growth in several geographically widely dispersed species of domesticated cotton resulted in similar and overlapping metabolic transformations of the manner in which cellular redox levels have become modulated.

Background

Reactive oxygen species (ROS), including singlet oxygen, superoxide anions, hydrogen peroxide (H_2O_2), and hydroxyl radicals, are generated by oxidative reactions and other metabolic processes in cells and could have deleterious effects on growth and survival [1-4]. Naturally, cells have evolved mechanisms to scavenge excess ROS to prevent cellular damage, including up-regulation of antioxidant defense mechanisms [5-8]. In addition to the necessity of controlling excess potentially damaging ROS, eukaryotes have harnessed ROS as signaling molecules for a diverse array of regulatory processes, including responses to abiotic and biotic stresses, regulation of growth and development, and control of programmed cell death [2-5,9-11]. Because of their important roles as signaling molecules, as well as their toxicity at higher levels, ROS concentrations are finely tuned and developmentally regulated by a complex gene network (at least 152 genes in *Arabidopsis*; [2]), which collectively control and modulate ROS metabolism [2,5].

Recently, several ROS-related signal transduction and sensing components have been identified, including kinases, calcium channel proteins and redox-sensitive transcription factors [12-17]. The role of ROS has also been studied in cell wall biosynthesis, where ROS have been shown to be involved in lignin biosynthesis, cross-linking reactions between cell wall components, and loosening of cell walls in growing tissues [18-22]. ROS have been proposed to be involved in regulating cell growth in root hairs and pollen tubes, for example, via NADPH oxidases that control development by generating ROS and regulate cell expansion through the activation of calcium channels [23]. The role of Ca^{2+} flux in ROS signaling is also well-characterized and involves calcium binding proteins, such as calmodulin and NADPH oxidases [24,25]. Stimulation of a Ca^{2+} influx into the cytoplasm through NADPH oxidase-derived ROS, and in turn, activation of NADPH oxidase to produce ROS also establish a positive feedback regulation maintaining growth in expanding root hair cells [26]. However, exogenous application of H_2O_2 attenuated the rate of root hair growth with a prolonged rise in Ca^{2+} after inhibition of growth [27]. ROS may also alter cell-wall properties and participate in their metabolism, as shown *in vivo* in radish seeds and maize roots [19,28]. In young cotton "fibers", which are single-celled epidermal trichomes, H_2O_2 appears to be important for the differentiation of the cellulose-rich secondary cell wall, and H_2O_2 levels are finely regulated [20,29]. Additionally, exogenous H_2O_2 levels are regulated by redox status-related antioxidant enzymes including Cu/Zn-superoxide dismutase (CSD) that localizes to secondary cell walls of developing cotton fibers and is involved in cell wall growth [22]. Redox levels in cotton fiber cells are important for stability of cellulose synthases, neces-

sary for cellulose biosynthesis during fiber elongation and secondary wall formation [30].

Since ROS levels are important to cell growth, the possibility exists that ROS modulation has been responsive to selection pressure. The genus *Gossypium* is an excellent model for studying the evolution of ROS modulation because cotton fiber represents a highly derived modification of epidermal seed trichomes. These trichomes have the experimental advantage of being single-celled and readily detached for *in vitro* studies of gene expression or for other purposes. From an evolutionary standpoint, trichome length, color and form vary considerably among the ~50 mostly wild species in the genus [31,32], thus providing a natural system in which to investigate the evolutionary relationships between cell growth and ROS modulation. Finally, *Gossypium* includes both diploids and allopolyploids, of which four species (two diploid and two allopolyploid) were independently domesticated for use as feed and fiber crops [32] (Figure 1). This evolutionary context, and especially the multiple, parallel replicated "experiments" of domestication over the past ~7000 years, provide a unique opportunity to explore the connections between cell growth, natural and human-mediated selection, and ROS modulation.

Initial insight into these relationships has emerged from recent comparative gene expression profiling experiments. In a comparison between a short-fibered, wild diploid species (*G. longicalyx*) and a domesticated diploid cotton (*G. herbaceum*), Hovav *et al.* [33] showed that during fiber development several stress responsive genes were up-regulated in the wild species, whereas ROS scavenging and signaling genes were comparatively over-expressed at the same developmental stages in the domesticated species. Analogous results were obtained in another microarray comparison in cotton, this time between wild and domesticated forms of the allopolyploid species *G. barbadense*. In this latter study, a greater number of antioxidant and ROS signaling genes were up-regulated in the domesticated accession early in fiber development, whereas at the same developmental stages genes associated with ROS production were over-expressed in the wild accession [34].

Here we build upon these earlier observations with new analyses and experiments to explore the expression of ROS-related genes during various evolutionary stages encompassing several orders of magnitude of differences in time, including ~5–10 millions of years of diploid species divergence, the diversification of wild allopolyploid species over the past 1–2 million years, and two parallel domestications of allopolyploid species approximately 5000 years ago [35,36]. We use high-resolution microarray methodology [37] to comparatively study gene expression for ~42,000 genes in diploid and allotetraploid

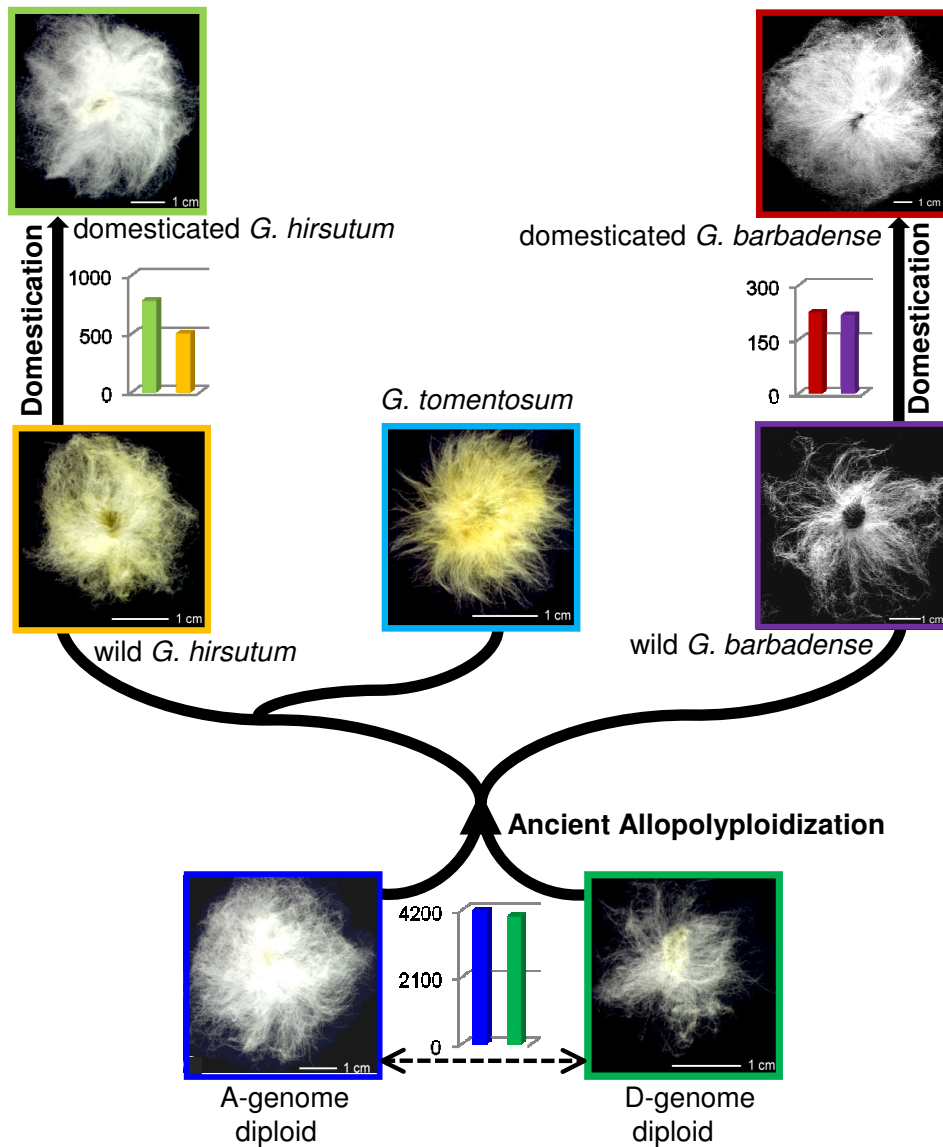


Figure 1

Evolutionary history of diploid and allotetraploid cotton (*Gossypium*). Phylogeny of the genus is shown, with the history of repeated domestication at both the diploid ($n = 13$) and polyploid ($n = 26$) levels. The ancestral A- and D-genome diploids are inferred to have diverged from a common ancestor ~5–10 million years ago, prior to genomic merger in a common polyploid nucleus ~1–2 million years ago [32]. The newly evolved allopolyploid lineage subsequently diversified into five species (three used for microarray analysis are shown). Two allotetraploid species, *G. hirsutum* (source of 'upland cotton') and *G. barbadense* (source of 'Pima cotton'), and the diploid species *G. arboreum* were domesticated by humans within the past ~7000 years [35,36]. In the present study, we used models of the two progenitor diploids, *G. arboreum* (A-genome) and *G. raimondii* (D-genome), and both wild and domesticated forms of *G. hirsutum* and *G. barbadense*. We also included *G. tomentosum*, a wild allopolyploid from Hawaii. A representative image of a single seed at maturation is also shown for each species, with attached trichomes ("cotton fiber") with average fiber length (*G. raimondii* = 1.6 cm; *G. arboreum* = 3.0 cm; wild *G. hirsutum* = 2.0 cm; domesticated *G. hirsutum* = 3.9 cm; wild *G. barbadense* = 2.4 cm; domesticated *G. barbadense* = 4.3 cm; *G. tomentosum* = 1.2 cm) (modified from Applequist *et al.* [31]). Arrows denote microarray comparisons between species, with graphs designating the number of differentially up-regulated genes ($P < 0.05$ and FDR < 0.01) in each species, using the same color codes as in the seed image outlines.

cotton species at an early stage of fiber development. By restricting our attention to ROS-related processes, we hoped to shed light on how ROS gene expression may have been altered in this single cell type during diploid divergence and natural allopolyploid formation, and by human selection pressure associated with repeated domestication.

Results and discussion

ROS-related gene expression evolution at the diploid level

To explore the evolution of oxidative stress related genes during diploid divergence, we assayed mRNA expression levels in fiber cells from domesticated *G. arboreum* and from wild *G. raimondii* at 2 days post-anthesis (hereafter "dpa"). This comparison encompasses natural species evolution in addition to human-mediated selection associated with domestication of the former species, which has spinnable fibers that are several-fold longer than the short, tightly adherent, commercially useless fibers of the former species (Figure 1). Our rationale for exploring fibers from 2 dpa was based on our earlier observations that oxidant and antioxidant genes are up- or down-regulated early in fiber development [33,34], but not later. For example, among the highest 100 differentially expressed genes in a wild accession of *G. barbadense* at 2 dpa, more than a third are potentially involved in stress-related processes; at later stages of fiber development, there were few such genes [34]. Using a custom microarray, this analysis revealed a high level of differential expression, using relatively stringent statistical criteria. Genes were considered significant if the *q* values, adjusted for multiple testing [38], were < 0.01 (Figure 1). Approximately 20% of all genes were differentially expressed between the two species, with a relatively even distribution in terms of direction; that is, 4,175 genes (~10% of the genes on the array) were up-regulated in *G. arboreum*, whereas 3,984 genes (~9.5%) were up-regulated in *G. raimondii*. This observation of extensive expression evolution between homologous cells of congeneric species, on a 5–10 million year timescale, is consistent with other recent comparative expression profiling experiments [33], where ~22% of all genes were differentially expressed in 5 dpa fibers of *G. longicalyx* and *G. herbaceum* species, consistent with the closer phylogenetic relationship of these taxa [32,39].

Among these more than 8,000 differentially expressed genes, gene lists were filtered for biological processes corresponding to putative ROS producing, scavenging and cell-signaling mechanisms based on GO annotations, as reviewed in [1] and [2] (Table 1), yielding a total of 548 genes (251 and 297 up-regulated in *G. arboreum* and *G. raimondii*, respectively; for gene list see Additional file 1). Notably, up-regulated genes were asymmetrically distributed between the two species with respect to classes of ROS genes. Many genes with antioxidant and transferase activity, and potential ROS-mediated cell-signaling proc-

esses are up-regulated in *G. arboreum* fibers, including 'peroxidase activity', 'glutathione transferase activity' and 'epoxide hydrolase activity', and in contrast few genes were categorized as contributing to ROS production. In contrast, *G. raimondii* fibers had higher levels of expression for genes involved in ROS production, including 'glycolate oxidase activity' and 'H₂O₂ biosynthetic process', but lower levels of expression for genes involved in ROS scavenging (Tables 1 & Additional file 2).

The foregoing results show parallels with earlier studies involving microarrays and cotton fiber development. Specifically, in comparisons between the diploid species *G. herbaceum* and *G. longicalyx*, as well as for wild vs. domesticated accessions of *G. barbadense* (Pima cotton), some genes involved with modulating H₂O₂ and other ROS levels were up-regulated in the accessions with longer fibers [33,34]. As noted above, inhibition of H₂O₂ production or application of exogenous H₂O₂ acts as a signal in cell wall differentiation in young cotton fibers, with extended cell elongation correlated with activation of antioxidant enzymes and premature formation of secondary walls being associated with increased H₂O₂ levels [20,29].

In addition to genes likely to be *directly* involved with ROS production and scavenging, many other genes may be stimulated through ROS signaling cascades that ultimately influence gene expression. For instance, although signaling molecules such as 'kinases' are known to be important in many cellular processes, some play roles in downstream signaling events associated with ROS. Serine/ threonine protein kinases for example, play a major role in ROS sensing and the activation of MAPKs [40]. However, downstream signaling events associated with ROS sensing involve calcium and calcium binding proteins [24,25]. In addition, a number of transcription factors are important components of the oxidative stress response signal transduction network. These include heat shock transcription factors, Zn finger proteins and WRKY transcription factors [12-15]. To search for the involvement of these classes of genes less certain to be involved in ROS metabolism than those directly functioning in the production and scavenging of ROS, we conducted searches based on sequence similarity to angiosperm homologs (usually from *Arabidopsis*), and identified several genes up-regulated in both *G. arboreum* and *G. raimondii* that become candidates for having roles in ROS signaling (Table 1). These, however, require experimental characterization to substantiate a role in ROS signaling.

Modulation of ROS levels may also be linked to cell elongation, as indicated by the aforementioned work on hydrogen peroxide levels and fiber elongation [20,29]. As shown in Table 1, more processes involving antioxidant genes (ROS scavengers) are up-regulated in *G. arboreum*

Differences between the MPV and the three wild polyploid species (*G. hirsutum*, *G. barbadense* and *G. tomentosum*) led to varying numbers of differential expression in each case (Figure 3), with 778 shared genes that were up-regulated. These shared genes were thus inferred to represent the early effects of polyploidization, prior to speciation and later domestication. Categorization of these genes into biological processes revealed no significantly over-represented process related to ROS metabolism (see Additional file 3), consistent with our expectation that genome merger and polyploidization would not affect ROS-related genes in the wild species. We explored the data further for genes up-regulated in each of three wild species as well as genes shared by any two of the three species (numbers shown in intersections in Figure 3). No process related to ROS metabolism emerged for any single species, whereas for pairs of species, there was a marginally significant up-regulation of 'oxidases' in wild *G. hirsutum* vs wild *G. barbadense* as well as wild *G. barbadense* vs *G. tomentosum* (see Additional file 4).

ROS-related genes and parallel domestication of cultivated allopolyploid cotton

We hypothesized that if ROS-related processes were sensitive to modification by selection via their putative rela-

tionship to cell elongation [33,34], then, and in contrast to the expectation for wild polyploids, domesticated varieties of the polyploid species would be expected to exhibit evidence of this at the gene expression level. Additionally, as noted above, two species of allopolyploid cotton, *G. hirsutum* and *G. barbadense* have been independently domesticated [32,35], thus allowing two tests of this hypothesis.

To test this we used the microarray data to perform wild/domesticated contrasts in both species. These analyses revealed 777 and 498 genes that were up- and down-regulated, respectively, in cultivated *G. hirsutum* versus its wild counterpart, and 223 and 216 that were up-and down-regulated, respectively, in cultivated *G. barbadense*, relative to its wild accession (Figure 1). Assessment of biological processes corresponding to these differentially up-regulated genes showed significant up-regulation of processes involving antioxidant genes in both domesticated species (see Additional file 2 & Table 2).

An important pattern emerged from these comparative expression data, namely, that shared processes between the two species were often observed but that they involved in different sets of genes. For example, the process 'perox-

Table 2: Biological processes over-represented (p < 0.05) in domesticated *G. hirsutum* and domesticated *G. barbadense* relative to their wild forms.

Domesticated species	GO term	
	up-regulated	down-regulated
<i>G. hirsutum</i>	antioxidant activity (Chl-APX, GST, CAT, TRX) glutathione transferase activity (GSTs) Glycoside hydrolase activity (GLH) peroxidase activity (APX, GST, CAT) glutathione S-conjugate-exporting ATPase activity removal of superoxide radicals (TRX) oxidoreductase activity, acting on peroxides (APX, GST, CAT) voltage-gated calcium channel activity (two pore Ca ⁺ channels)	regulation of O ₂ ⁻ /H ₂ O ₂ production process (NADPH oxidase) glycolate oxidase activity (GOX) cyclin-dependent protein kinase inhibitor activity jasmonic acid and ethylene-dependent systemic resistance
<i>G. barbadense</i>	response to stress (ESP, PRX, GH3-like) peroxidase activity (PRX) antioxidant activity (Chl-APX, PRX) regulation of epoxide hydrolase activity (EPH) voltage-gated calcium channel activity (two pore Ca ⁺ channels) MAP kinase activity (MAPK1, WIPK) receptor signaling protein serine/threonine kinase activity (MAK, MAPKs) jasmonic acid mediated signaling pathway (GH3-like) glycolate oxidase activity (GOX)	superoxide dismutase activity (Cu/Zn SOD) carotenoid biosynthetic process (GPS) ent-kaurene oxidase activity (CYP450) oxygen and reactive oxygen species metabolic process (SOD) photosystem II reaction center

Key genes are shown in parentheses (a complete list can be viewed in Additional file 5). Abbreviations: Chl-APX – Thylakoid-bound ascorbate peroxidase; GST – Glutathione S-transferase; CAT – Catalase, TRX – Thioredoxin reductase; GLH – Glycoside hydrolase; APX – Ascorbate peroxidase; PRX – Peroxidase; GOX – Glycolate oxidase; EPH – Epoxide hydrolase; MAPK – Mitogen-activated protein kinase; SOD – Superoxide dismutase; ESP – Ethylene signaling protein; GPS – geranylgeranyl pyrophosphate synthase.

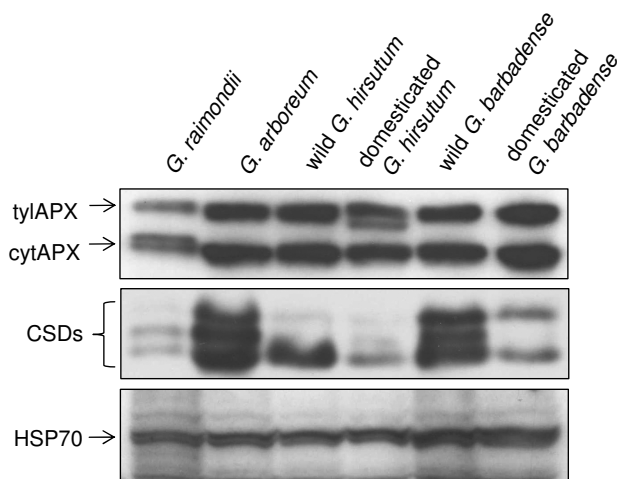


Figure 2
Protein blot analysis from wild and domesticated diploid and polyploid species. It is to show levels of thylakoid bound and cytosolic ascorbate peroxidase (tyLAPX and cytAPX), Cu/Zn-Superoxide dismutase (CSD) and HSP70 proteins in 2 dpa fiber extracts. All experiments were repeated twice and representative results are shown.

idase activity' in *G. hirsutum* includes ascorbate peroxidase, catalase and thioredoxin reductase genes, whereas in *G. barbadense* this process is represented by several peroxidase genes (see Additional file 2 & 4). Similarly, different sets of genes in the 'antioxidant activity' category were up-regulated in the two species, i.e., catalase, glutathione S-transferases and thioredoxin in *G. hirsutum*, and peroxidases in *G. barbadense* (Table 2 & Additional file 5). These genes have been shown to function as antioxidants, assisting in the regulation of ROS levels and contributing to cell elongation under high redox conditions [8,29,41,42]. Our data suggest that parallel selection by humans operated on different genetic components of the fiber development program in *G. hirsutum* and *G. barbadense*, leading to parallelism at the metabolic level and in morphology.

In addition to parallel up-regulation of ROS-related processes, our data also reveal possible examples of parallel down-regulation. A case in point is for the process 'peroxide biosynthesis', which was down-regulated in the cultivated varieties of both *G. hirsutum* and *G. barbadense*, suggesting that down-regulation of ROS production accompanied domestication (Table 2). In addition to the foregoing examples of putative parallel responses to domestication, other ROS-related processes and genes were significantly differentially expressed in one of the two domesticated species. In domesticated *G. hirsutum*, the biological process 'cyclin-dependent protein kinase inhibitor activity', which harbors a gene for inhibiting serine/threonine kinase that has been shown to play crucial role in ROS sensing [40], was down-regulated. This obser-

vation raises the possibility that domestication entailed an repression of kinase inhibitor genes, enhancing the potential of a growing cell to sense toxic ROS levels and activate downstream processes to maintain optimal concentration. In domesticated *G. barbadense*, the processes 'superoxide dismutase' and 'carotenoid biosynthetic process' were down-regulated, both known to be important in ROS scavenging (Table 2) [6,41]. The microarray based interpretations were also confirmed with comparable results at protein level for Cu/Zn-superoxide dismutase (CSD) and tyLAPX and cytAPX for wild and domesticated *G. hirsutum* and *G. barbadense* (Figure 2). Comparable to microarray data, higher levels of CSD is observed in wild *G. barbadense* in comparison to domesticated *G. barbadense* (Figure 2). However, although CSDs transcript levels in domesticated *G. hirsutum* are up-regulated in comparison to wild *G. hirsutum* (see Additional file 2), this is not observed in the protein blots (Figure 2). This inconsistency could be a result of antibodies cross-reacting with only some but not all of the CSD gene products, potentially not reacting with the specific CSD transcripts up-regulated in domesticated *G. hirsutum*, as well as differences in post-translational modification, or other factors.

ROS-related genes and parallel domestication across ploidy levels and continents

Because cotton domestication involved in two different allopolyploid species in the New World as well as two different diploid species in the Old World [32,36], a naturally replicated "experiment" of domestication provides the opportunity to evaluate potential parallelisms in ROS-related responses across ploidy levels and continents. To explore parallel recruitment of genes during domestication at the diploid and polyploid levels, we tabulated the differentially expressed genes (up- and down-regulated) that were shared between *G. arboreum* and domesticated *G. hirsutum* as well as between *G. arboreum* and domesticated *G. barbadense* (marked by asterisks in Additional file 2). A total of 178 and 38 shared genes were up-regulated and 102 and 22 were down-regulated in *G. arboreum* with *G. hirsutum* and *G. arboreum* with *G. barbadense* in comparisons to their wild relatives, respectively. Up-regulated genes shared by *G. arboreum* and *G. hirsutum* belong mainly to ROS scavenging classes, including glutathione S-transferase and ascorbate peroxidases (see Additional file 2), but also include putative ROS signaling genes such as serine/threonine kinase (see Additional file 5). Up-regulation in *G. arboreum* and *G. hirsutum* suggests that selection during domestication for agronomically more favorable fiber phenotypes resulted, in parallel, in the enhanced expression of similar genes. In contrast, *G. raimondii* and wild *G. hirsutum* have low levels of expression of these same genes, correlated with their much shorter fibers and temporally compressed development (Figure 4a) [31].

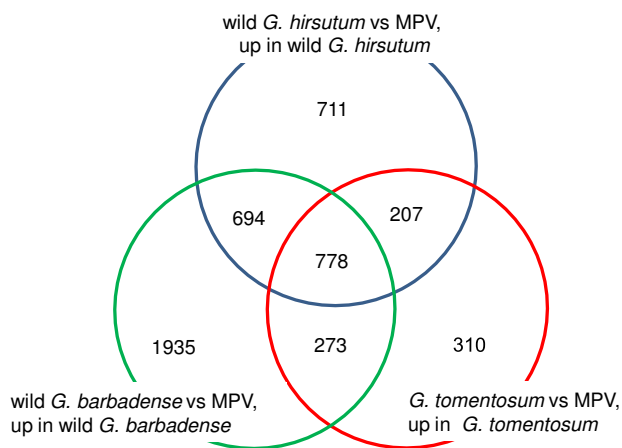


Figure 3
Venn diagrams of gene expression evolution in allopolyploid cotton. The numbers indicate genes classified as up-regulated in any wild polyploid species in comparison to the mid-parent values (MPV; estimated from a 1:1 RNA mix from the diploids *G. arboreum* and *G. raimondii*). Intersections denote genes shared by two or more of the three wild forms of the allopolyploids (wild *G. hirsutum*, wild *G. barbadense* and *G. tomentosum*).

Shared up-regulated genes between *G. arboreum* and domesticated *G. barbadense* include a suite of peroxidase genes (see Additional file 2). These genes encode an important class of ROS-regulating enzymes, which eliminate toxic molecules generated as a byproduct of aerobic respiration [29]. As shown in Figure 4b, expression levels of peroxidase genes in domesticated *G. barbadense* were similar to those of *G. arboreum*, suggesting once again parallel selection under domestication at the diploid and polyploid levels.

Conclusion

The present study implicates an oxidative stress responsive gene network as being involved in the evolution of elongated epidermal seed trichomes, providing the foundation for later human domestication of an important crop plant. We provide clues here into ROS related genes that may have been selected by humans, starting with initial domestication of wild perennial thousands of years ago, through the development of modern elite lines. Notably, the majority of up-regulated ROS-related processes are diagnosed as having become enhanced during domestication, at both the diploid and polyploid levels, as opposed to being a result of allopolyploid formation per se. This suggestion that expression evolution of oxidative stress related genes was primarily concomitant with domestication is bolstered by the remarkable observation that independent parallel domestication events, taking place in different hemispheres thousands of years ago under aboriginal human selection pressure, led to parallel recruit-

ment of ROS scavenging and signaling genes in one diploid and two different polyploid species. Although this is true at the level of biological process, or perhaps metabolism, it is clear that the genesis of these similarities is only partially congruent at the genetic level. That is, different sets of antioxidant genes are up-regulated in the domesticates. This repeated metabolic transformation accompanying domestication would appear to be without precedent. An exciting prospect for future work will be to dissect this physiological parallelism into its responsible constituent genes, and to learn the mutational basis of their altered regulation or function. Notwithstanding our ignorance of the specific function of most of the genes implicated as important in this study, a generality emerges that the avoidance or delay of stress-like processes in domesticated species may play a large role in the enhance elongation of fibers in cultivated compared to wild forms, in conjunction with an increased ability to modulate cellular redox balance in the growing cell. It would be interesting to explore whether similar trends in the evolution of ROS gene expression accompanied the domestication of other crops and for other traits, for example tolerance to stressed environmental conditions or to enhanced growth of other organs.

Methods

Plant materials, RNA isolation and microarray preparation

Three biological replicates of seven *Gossypium* accessions were grown in the Pohl Conservatory at Iowa State University, Ames, IA: *G. arboreum*, *G. raimondii*, a wild form of *G. hirsutum* (accession TX2094), a domesticated form of *G. hirsutum* (accession TM1), a wild form of *G. barbadense* (accession K101), a domesticated form of *G. barbadense* (accession Pima S-7) and *G. tomentosum*. Because truly wild, as opposed to feral, forms of *G. barbadense* are difficult to verify, we selected the accession used based on earlier allozyme analysis (Percy and Wendel, 1990) and its relatively primitive morphology. These accessions include representatives of both diploid progenitor genomes ("A" and "D") of allopolyploid cotton [32](Figure 1), and both wild and domesticated forms of allopolyploid cotton (*G. hirsutum* and *G. barbadense*).

Flowers were tagged at anthesis and harvested at 2 days post-anthesis. For each of three biological replicates, ovules were excised, frozen in liquid nitrogen, and stored at -80°C. RNA extractions were performed following a modified hot borate procedure optimized for *Gossypium* [43]. From each pair of A₂ and D₃ replicates, an equimolar RNA mix (1:1 mix) was also generated to estimate mid-parent expression values (MPV). RNA samples were amplified with the TargetAmp™ 1-Round aRNA Amplification kit from Epicentre Biotechnologies (Madison, WI) and quantified and assessed for degradation using a Bioanalyzer (Agilent Technologies, Santa Clara, CA). All

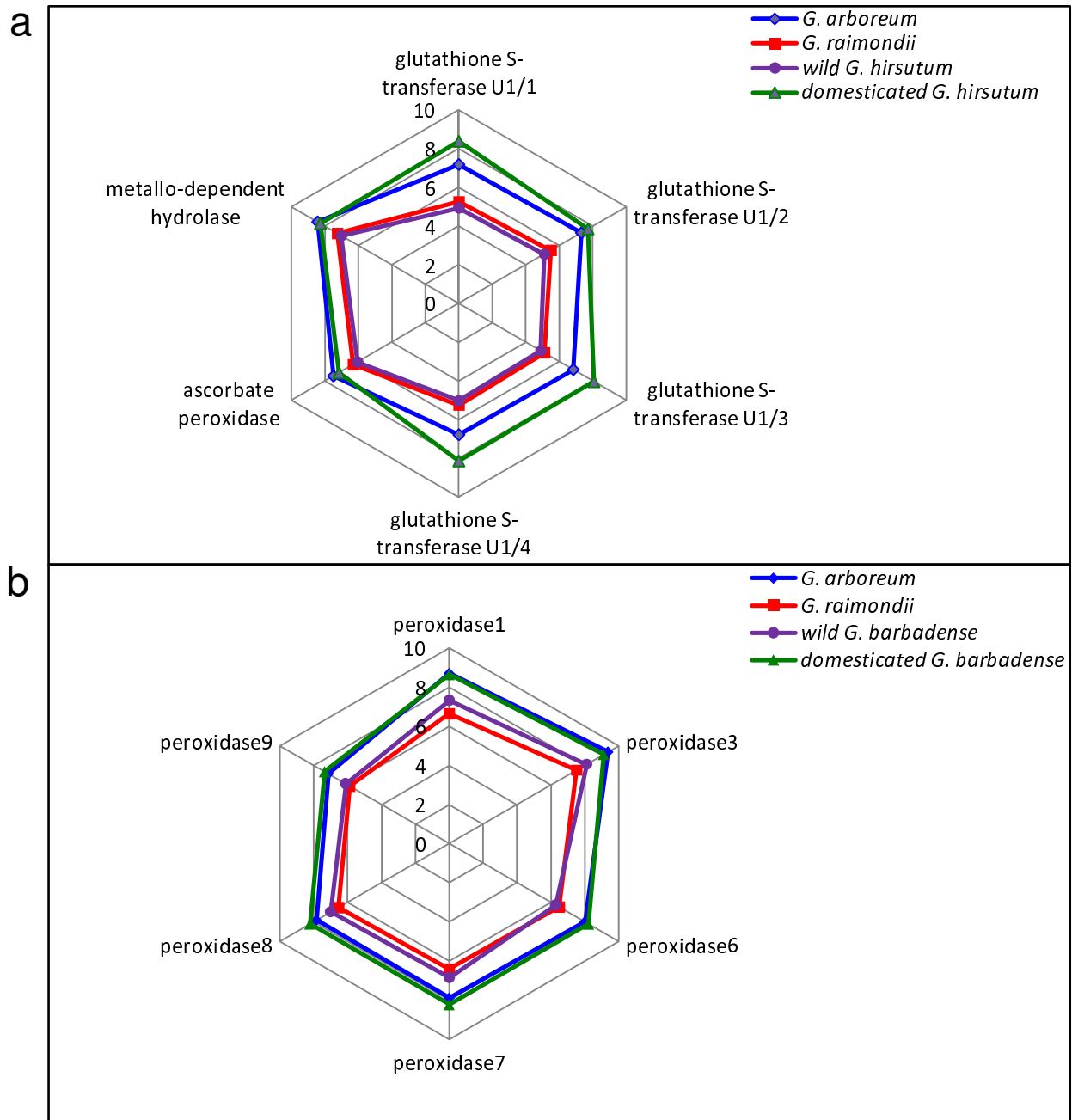


Figure 4
Differential gene expression patterns for some ROS-related genes. These genes are differentially up-regulated in *G. arboreum* and the domesticated polyploids in contrast to their relative wild relatives or ancestors. Each point on the polygons represents a gene and the scale corresponds to relative expression levels in the diploid model progenitor genomes (*G. arboreum* and *G. raimondii*) and the wild and domesticated forms of: (a) *G. hirsutum*; and (b) *G. barbadense*.

amplified RNA samples were sent to NimbleGen Systems (Madison, WI) for cDNA synthesis, labeling, and hybridization to custom microarrays developed from an assembly of cotton ESTs [37]. This custom microarray platform has the ability to measure homoeolog-specific gene expression for ~1500 genes in the allotetraploids, as well as overall gene expression for 42,429 unigenes.

Microarray data processing and statistical analysis

Raw data values for each gene were natural-log transformed, median-centered, and quantile-normalized across all arrays. Following normalization, contig-level log ratio values were determined by calculating an average of log ratio values from 7 probes per gene for a total of 42,429 genes, using Tukey's biweight method [44,45]. A linear model as described [34], including effects for replication and genotype, was fitted to these contig-level data, allowing the estimation of all possible contrasts between *G. arboreum*, *G. raimondii*, their 1:1 mix, wild *G. hirsutum*, domesticated *G. hirsutum*, wild *G. barbadense*, domesticated *G. barbadense* and *G. tomentosum*. For each gene, differences were calculated using pair-wise contrasts between A₂ vs D₅, wild *G. hirsutum* vs 1:1 mix, wild *G. barbadense* vs 1:1 mix, *G. tomentosum* vs 1:1 mix, wild *G. hirsutum* vs domesticated *G. hirsutum* and wild *G. barbadense* vs domesticated *G. barbadense*. The 42,429 *p*-values from each comparison were converted to *q*-values using the method of [38]. These *q*-values were used to identify the number of differentially expressed genes for a given comparison when controlling for a false discovery rate (FDR) of 0.01. Blast2GO <http://www.blast2go.de> was used to infer biochemical pathways involved in a given comparison and to calculate statistical significance. The expression values for microarray experiments have been submitted to the database with following accession numbers: GSM432872; GSM432873; GSM432874; GSM432875; GSM432876; GSM432877; GSM432878; GSM432879; GSM432880; GSM432881; GSM432882; GSM432883; GSM432884; GSM432885; GSM432886; GSM432887; GSM432888; GSM432889.

Validation of microarray results with massARRAY

To confirm the microarray-based interpretations, we used an application of a mass ARRAY based validation technique. The robustness of mass ARRAY technique has been demonstrated showing high correlation with RT-PCR [46]. We checked 18 randomly selected genes at 2 dpa in three biological replicates for homoeolog contributions to the transcriptome for domesticated *G. hirsutum* samples (for details of validation design, see Supplementary methods in Additional file 6). The mean value for each of three replicates was determined and compared with the estimates derived from the homoeolog-specific microarray. The correlation between SNP-specific microarray and

mass ARRAY-based techniques is shown with high R² value (0.84) at *p*-value < 0.001 in Additional file 6.

Protein blot analysis

Cotton fibers were mechanically separated from frozen ovular tissue using a modified liquid nitrogen and glass bead shearing procedure [47]. Total protein samples were extracted using a phenol-ammonium acetate/methanol method [48]. Briefly, ~250 mg of crushed fiber tissue was dissolved in extraction buffer (100 mM Tris-HCl, (pH 8.8), 10 mM EDTA, 900 mM sucrose, 0.4% 2-mercaptoethanol) at 4°C. The proteins were precipitated overnight with ammonium acetate/methanol solution at -20°C and the pellet, collected by centrifugation, was washed with ice-cold ammonium acetate/methanol (100 mM ammonium acetate in 100% methanol) and 80% acetone and dried. The total proteins pellets were solubilized into 1× Laemmli sample buffer (0.0625 M Tris-HCl (pH 6.8), 2% SDS, 25% glycerol, 5% 2-mercaptoethanol) and stored at -20°C until analysis. Protein gel blots were performed as described [7]. Antibodies against cytosolic and chloroplastic APXs (ascorbate peroxidase) were obtained using a domain conserved to these proteins (a fragment of thylakoid APX from Lys¹⁰⁰ to Ile³⁴¹) as described in [49]. Antibodies against Cu/Zn-superoxide dismutase (CSD) was obtained from Agrisera (Vannas, Sweden) and that against HSP70 was prepared as described by [50] using purified proteins as antigens. Primary rabbit antibodies were used in a 1:3000 dilution while secondary goat anti rabbit antibodies conjugated to horse radish peroxidase (Invitrogen, Carlsbad, CA) were used at a 1:20,000 dilution. ECL™ chemiluminescent detection (Amersham Biosciences, Buckinghamshire, UK) was performed as described in [7].

Authors' contributions

BC and JFW designed the research work and wrote the manuscript. BC, RH and RM carried out the laboratory-based studies. BC and LF analysed the data. RM and JFW advised on numerous aspects of the study, reviewed the results and performed critical reading and editing of the manuscript. All authors read and approved the final manuscript.

Additional material

Additional file 1

Biological processes with their corresponding genes up-regulated in G. arboreum in contrast to G. raimondii. The data provided represent the statistical comparison of two diploid species at early stage of fiber development.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-10-378-S1.xls>]

Additional file 2

Gene annotation and significant over-representation of the ROS-scavenging network in cotton fibers (FDR < 0.01). The data provided represents all up- or down-regulated antioxidant genes in domesticated accession in comparison to their wild relatives.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-10-378-S2.xls>]

Additional file 3

*Common biological processes up- and down-regulated after polyploid formation in three wild allopolyploid species (wild *G. hirsutum*, *G. tomentosum* and wild *G. barbadense*) in contrast to the mid-parent value (MPV) from progenitor diploid genomes. The data provided represents the up-regulated genes in all three wild species after polyploidization.*

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-10-378-S3.doc>]

Additional file 4

Biological processes categorized based on differentially up-regulated unique genes in each of three wild polyploid species in contrast to the MPV, exclusive of any set of shared genes; and processes based on up-regulated genes shared by any two species (exclusive of up-regulated genes shared by all three wild species). The data provided represents the unique and shared biological processes observed to be up-regulated only in all three wild accessions studied.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-10-378-S4.doc>]

Additional file 5

Assessment of biological processes for up-regulated genes in domesticated accessions relative to their wild forms. The data provided represents all significant biological process observed to be up-regulated in two domesticated accession in comparison to their wild relatives.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-10-378-S5.xls>]

Additional file 6

Validation of NimbleGen microarray values by mass ARRAY-based homoeolog-specific expression measurements. (a) List of 18 homoeologous genes analyzed for validation of NimbleGen microarray values by mass ARRAY-based homoeolog-specific expression measurements. (b) Correlation between NimbleGen microarray data and mass ARRAY-based homoeolog-specific measurements. The relative expression of D-homoeolog detected at 2 dpa based on NimbleGen microarray signal information and plotted on the y-axis. The x-axis is the proportion of D-homoeolog that is observed based on mass ARRAY measurements. The best-fit trend line with R² correlation and p-value is shown on the graph for early stage of fiber development. The data provided represents the validation of homoeologous gene expression in the developing fibers from wild and domesticated accessions.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-10-378-S6.pdf>]

Acknowledgements

We thank Guanqing Hu and Luhua Song for their help in protein isolation and western blotting. This project was supported by the United States National Science Foundation Plant Genome Program, by IOS-0743954, and a fellowship from the Department of Biotechnology, Government of India to BC. RM would like to thank ISF grant 214/08, as well as EU FP7 – MARIE CURIE 447.

References

- Mittler R: **Oxidative stress, antioxidants and stress tolerance.** *Trends Plant Sci* 2002, **7(9)**:405-410.
- Mittler R, Vanderauwera S, Gollery M, Van Breusegem F: **Reactive oxygen gene network of plants.** *Trends Plant Sci* 2004, **9(10)**:490-498.
- Mitsuhashi I, Malik KA, Miura M, Ohashi Y: **Animal cell-death suppressors Bcl-xL and Ced-9 inhibit cell death in tobacco plants.** *Curr Biol* 1999, **9**:775-778.
- Dat J, Vandenaabeele S, Vranová E, Van Montagu M, Inzé D, Van Breusegem F: **Dual action of the active oxygen species during plant stress responses.** *Cell Mol Life Sci* 2000, **57**:779-795.
- Apel K, Hirt H: **Reactive oxygen species: Metabolism, oxidative stress, and signal transduction.** *Ann Rev Plant Biol* 2004, **55(1)**:373-399.
- Bowler C, Montagu MV, Inze D: **Superoxide dismutase and stress tolerance.** *Ann Rev Plant Physiol Plant Mol Biol* 1992, **43(1)**:83-116.
- Davletova S, Rizhsky L, Liang H, Shengqiang Z, Oliver DJ, Coutu J, Shulaev V, Schlauch K, Mittler R: **Cytosolic ascorbate peroxidase I is a central component of the reactive oxygen gene network of Arabidopsis.** *Plant Cell* 2005, **17**:268-281.
- Willekens H, Chamnongpol S, Davey M, Schraudner M, Langebartels C, Montagu MV, Inze D, Camp WV: **Catalase is a sink for H₂O₂ and is indispensable for stress defence in C₃ plants.** *EMBO J* 1997, **16**:4806-4816.
- Lopez-Huertas E, Charlton WL, Johnson B, Graham IA, Baker A: **Stress induces peroxisome biogenesis genes.** *EMBO J* 2000, **19**:6770-6777.
- Nakagami H, Pitzschke A, Hirt H: **Emerging MAP kinase pathways in plant stress signaling.** *Trends Plant Sci* 2005, **10**:339-346.
- Overmyer K, Brosché M, Kangasjärvi J: **Reactive oxygen species and hormonal control of cell death.** *Trends Plant Sci* 2003, **8**:335-342.
- Panchuk II, Volkov RA, Schoffl F: **Heat stress- and heat shock transcription factor-dependent expression and activity of ascorbate peroxidase in Arabidopsis.** *Plant Physiol* 2002, **129(2)**:838-853.
- Pnueli L, Liang H, Rozenberg M, Mittler R: **Growth suppression, altered stomatal responses, and augmented induction of heat shock proteins in cytosolic ascorbate peroxidase (Apx1)-deficient Arabidopsis plants.** *Plant J* 2003, **34(2)**:187-203.
- Rizhsky L, Davletova S, Liang H, Mittler R: **The zinc finger protein Zat12 is required for cytosolic ascorbate peroxidase expression during oxidative stress in Arabidopsis.** *J Biol Chem* 2004, **279(12)**:11736-11743.
- Rizhsky L, Liang H, Mittler R: **The water-water cycle is essential for chloroplast protection in the absence of stress.** *J Biol Chem* 2003, **278(40)**:38921-38925.
- Vandenbroucke K, Robbens S, Vandepoele K, Inze D, Peer Y Van de, Van Breusegem F: **Hydrogen peroxide-induced gene expression across kingdoms: A comparative analysis.** *Mol Biol Evol* 2008, **25(3)**:507-516.
- Vanderauwera S, Zimmermann P, Rombauts S, Vandenaabeele S, Langebartels C, Gruijssem W, Inze D, Van Breusegem F: **Genome-wide analysis of hydrogen peroxide-regulated gene expression in Arabidopsis reveals a high light-induced transcriptional cluster involved in anthocyanin biosynthesis.** *Plant Physiol* 2005, **139**:806-821.
- Fry SC: **Oxidative scission of plant cell wall polysaccharides by ascorbate-induced hydroxyl radicals.** *Biochem J* 1998, **332**:507-515.

19. Liszkay A, Zalm E van der, Schopfer P: **Production of reactive oxygen intermediates (O_2^- , H_2O_2 , and OH) by maize roots and their role in wall loosening and elongation growth.** *Plant Physiol* 2004, **136**:3114-3123.
20. Potikha TS, Collins CC, Johnson DI, Delmer DP, Levine A: **The involvement of hydrogen peroxide in the differentiation of secondary walls in cotton fibers.** *Plant Physiol* 1999, **119**:849-858.
21. Pomar F, Caballero N, Pedreño MA, Ros Barceló A: **H_2O_2 generation during the auto-oxidation of coniferyl alcohol drives the oxidase activity of a highly conserved class III peroxidase involved in lignin biosynthesis.** *FEBS Letters* 2002, **529**:198-202.
22. Kim H, Kato N, Kim S, Triplett B: **Cu/Zn superoxide dismutases in developing cotton fibers: evidence for an extracellular form.** *Planta* 2008, **228**(2):281-292.
23. Foreman J, Demidchik V, Bothwell JHF, Mylona P, Miedema H, Torres MA, Linstead P, Costa S, Brownlee C, Jones JDG, et al.: **Reactive oxygen species produced by NADPH oxidase regulate plant cell growth.** *Nature* 2003, **422**(6930):442-446.
24. Bowler C, Fluhr R: **The role of calcium and activated oxygens as signals for controlling cross-tolerance.** *Trends Plant Sci* 2000, **5**(6):241-246.
25. Knight H, Knight MR: **Abiotic stress signalling pathways: specificity and cross-talk.** *Trends Plant Sci* 2001, **6**(6):262-267.
26. Takeda S, Gapper C, Kaya H, Bell E, Kuchitsu K, Dolan L: **Local positive feedback regulation determines cell shape in root hair cells.** *Science* 2008, **319**(5867):1241-1244.
27. Jones DL, Gilroy S, Larsen PB, Howell SH, Kochian LV: **Effect of aluminum on cytoplasmic Ca^{2+} homeostasis in root hairs of *Arabidopsis thaliana* (L.).** *Planta* 1998, **206**(3):378-387.
28. Schopfer P, Plachy C, Frahrig G: **Release of reactive oxygen intermediates (superoxide radicals, hydrogen peroxide, and hydroxyl radicals) and peroxidase in germinating radish seeds controlled by light, gibberellin, and abscisic acid.** *Plant Physiol* 2001, **125**(4):1591-1602.
29. Li H-B, Qin YM, Yu P, Wen-Qiang S, Wen-Qian M, Yu-Xian Z: **A cotton ascorbate peroxidase is involved in hydrogen peroxide homeostasis during fibre cell development.** *New Phytol* 2007, **175**:462-471.
30. Jacob-Wilk D, Kurek I, Hogan P, Delmer DP: **The cotton fiber zinc-binding domain of cellulose synthase A1 from *Gossypium hirsutum* displays rapid turnover in vitro and in vivo.** *Proc Natl Acad Sci USA* 2006, **103**:12191-12196.
31. Applequist WL, Cronn R, Wendel JF: **Comparative development of fiber in wild and cultivated cotton.** *Evol Devel* 2001, **3**:3-17.
32. Wendel JF, Cronn RC: **Polyploidy and the evolutionary history of cotton.** *Adv Agron* 2003, **78**:139-186.
33. Hovav R, Udall JA, Chaudhary B, Hovav E, Fligel L, Hu G, Wendel JF: **The evolution of spinable cotton fiber entailed natural selection for prolonged development and a novel metabolism.** *PLoS Genetics* 2008, **4**(2):e25.
34. Chaudhary B, Hovav R, Rapp R, Verma N, Udall JA, Wendel JF: **Global analysis of gene expression in cotton fibers from wild and domesticated *Gossypium barbadense*.** *Evol Devel* 2008, **10**(5):567-582.
35. Dillehay TD, Rossen J, Andres TC, Williams DE: **Preceramic adoption of peanut, squash, and cotton in northern Peru.** *Science* 2007, **316**:1890-1893.
36. Wendel JF: **Cotton.** In *Evolution of crop plants* Edited by: Simmonds N, Smartt J. London: Longman; 1995:358-366.
37. Udall JA, Fligel LE, Cheung F, Woodward AW, Hovav R, Rapp RA, Swanson JM, Lee JJ, Gingle AR, Nettleton D, et al.: **Spotted cotton oligonucleotide microarrays for gene expression analysis.** *BMC Genomics* 2007, **8**:81.
38. Benjamini Y, Hochberg Y: **Controlling the false positive discovery rate: A practical and powerful approach to multiple testing.** *J R Stat Soc* 1995, **57**:289-300.
39. Cronn RC, Small RL, Haselkorn T, Wendel JF: **Rapid diversification of the cotton genus (*Gossypium*: Malvaceae) revealed by analysis of sixteen nuclear and chloroplast genes.** *Am J Bot* 2002, **89**:707-725.
40. Rentel MC, Lecourieux D, Ouaked F, Usher SL, Petersen L, Okamoto H, Knight H, Peck SC, Grierson CS, Hirt H, et al.: **OXII kinase is necessary for oxidative burst-mediated signalling in *Arabidopsis*.** *Nature* 2004, **427**(6977):858-861.
41. Asada K, Takahashi M: **Production and scavenging of active oxygen in photosynthesis.** In *Photoinhibition* Edited by: Kyle DJ, Osmond CB, Arntzen CJ. Amsterdam: Elsevier; 1987:227-287.
42. Edwards R, Dixon DP, Walbot V: **Plant glutathione S-transferases: enzymes with multiple functions in sickness and in health.** *Trends Plant Sci* 2000, **5**(5):193-198.
43. Wan C, Wilkins T: **A modified hot borate method significantly enhances the yield of high-quality RNA from cotton (*Gossypium hirsutum* L.).** *Annal Biochem* 1994, **223**:7-12.
44. Tukey JW: **Exploratory data analysis.** Addison-Wesley, Reading, MA; 1977.
45. Velleman PF, Hoaglin DC: **The ABC of exploratory data analysis.** Duxbury Press, Belmont, CA; 1981.
46. Stupar RM, Springer NM: **Cis-transcriptional variation in maize inbred lines B73 and Mo17 leads to additive expression patterns in the F1 hybrid.** *Genetics* 2006, **173**:2199-2210.
47. Taliencio EW, Boykin D: **Analysis of gene expression in cotton fiber initials.** *BMC Plant Biol* 2007, **7**:22.
48. Agrawal GK, Thelen JJ: **A high-resolution two dimensional gel- and Pro-Q DPS-based proteomics workflow for phosphoprotein identification and quantitative profiling.** In *Methods in Molecular Biology Volume 527*. Edited by: Graauw Md. New Jersey, USA: The Humana Press; 2008:3-19.
49. Miller G, Suzuki N, Rizhsky L, Hegie A, Koussevitzky S, Mittler R: **Double mutants deficient in cytosolic and thylakoid ascorbate peroxidase reveal a complex mode of interaction between reactive oxygen species, plant development, and response to abiotic stresses.** *Plant Physiol* 2007, **144**(4):1777-1785.
50. Mittler R, Merquiol E, Hallak-Herr E, Rachmilevitch S, Kaplan A, Cohen M: **Living under a 'dormant' canopy: a molecular acclimation mechanism of the desert plant *Retama raetam*.** *Plant J* 2001, **25**(4):407-416.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

